

IMMUNOFLOW™

MYCOPLASMA PNEUMONIAE IGM TEST

IVD For In Vitro Diagnostic Use



INTENDED USE

ImmunoFLOW *Mycoplasma pneumoniae* IgM Test detects human IgM to the complement-fixing antigen and is intended as an aid in the diagnosis of infection.

SUMMARY

The order Mycoplasmatales includes approximately 70 species, most of which are not found in humans. The genus *Mycoplasma* contains two species commonly found in man, *M. pneumoniae* and *M. genitalium*. These two species share lipid antigen specificities (complement-fixing antigen), and are therefore antigenically related. Two other human pathogens, *M. hominis* and *Ureaplasma urealyticum* are not serologically related and therefore not cross-reactive to this complement-fixing antigen.

M. pneumoniae is the only known *Mycoplasma* species that is a primary pathogen in man. Clinical manifestations can range from asymptomatic respiratory infections to severe pneumonia (1). *M. pneumoniae* accounts for 15 to 20% of total pneumonia (2), (3). Other symptoms associated with *M. pneumoniae* infection include abnormalities of the central nervous system (meningitis, encephalitis), cardiac involvement (myocarditis, pericarditis), hemolytic anemia, arthritis, G.I. inflammations, and mucocutaneous reactions (4). *M. pneumoniae* is identified as a common infectious cause of Stevens-Johnson Syndrome, a well-defined systemic disease that can develop into a life-threatening illness in children (5).

The *M. pneumoniae* organism is sensitive to erythromycin and tetracyclines; however, it is resistant to drugs more routinely given in the treatment of acute pneumonia. Thus, a rapid and reliable diagnosis of *M. pneumoniae* infection is essential to proper patient management (6). *M. pneumoniae* culture is difficult, slow and relatively insensitive. Nucleic acid detection methods can be sensitive but require localized specimen collection (e.g., lower lung lavage). Serology is the primary diagnostic tool. Serological methods are complement fixation (CF), indirect immunofluorescence assays (IFA), immune adherence hemagglutination assay (IAHA) and enzyme immunosorbent assays (EIA).

ASSAY PRINCIPLE

ImmunoFLOW is an immunoassay consisting of a cassette and three reagents. The cassette contains a paper matrix (for example, nitrocellulose) and an absorbent material. The paper matrix was manufactured with three "dots", each contain an antigen (e.g., positive control, analyte 1, analyte 2). A body fluid (e.g., serum) is applied to the triangular opening and allowed to flow

through the paper matrix into the absorbent. To assure assay specificity, a wash reagent is applied and flows into the absorbent material. Finally, gold particles attached to an immunological reagent (e.g., anti-immunoglobulin) is applied and absorbed.

The "dot" applied to the paper matrix contains antigen. Specific antibody in body fluid will bind to the antigen. If specific antibody binding occurs, immunologically active gold particles will bind and cause a red/pink color formation.

REAGENTS

Cassette: *M. pneumoniae*, strain FH (ATCC #15531), complement-fixing antigen and reagent control

Wash 2: Buffered solution with <0.1% sodium azide

Diluent: Buffered saline with <0.1% sodium azide

Color M: Colloidal gold conjugated to goat anti-human IgM in buffered saline with <0.1% sodium azide

WARNINGS AND PRECAUTIONS

For In Vitro Diagnostic Use: ImmunoFLOW reagents have been optimized for use as a system. Do not substitute other manufacturers' reagents or other ImmunoFLOW Test reagents. Dilution or adulteration of these reagents may also affect the performance of the test. Do not use any kits beyond the stated expiration date. Close adherence to the test procedure will assure optimal performance. Do not shorten or lengthen stated incubation times since this may result in poor assay performance.

Sodium azide may react with lead and copper plumbing to form highly explosive metal azides. It may be harmful if enough is ingested (more than supplied in kit). On disposal of liquids, flush with a large volume of water to prevent azide build-up (7). This dilution is not subject to GHS, US HCS and EU Regulation 2008/1272/EC labeling requirements.



Human source material. Material used in the preparation of this product has been tested and found non-reactive for hepatitis B surface antigen (HBsAg), antibodies to hepatitis C virus (HCV), and antibodies to human immunodeficiency virus (HIV-1 and HIV-2). Because no known test method can offer complete assurance that infectious agents are absent, handle reagents and patient samples as if capable of transmitting infectious disease (8). Follow recommended Universal Precautions for bloodborne pathogens as defined by OSHA (9), Biosafety Level 2 guidelines from the current CDC/NIH Biosafety in Microbiological and Biomedical Laboratories (10), WHO Laboratory Biosafety Manual (11), and/or local, regional and national regulations.

STORAGE

Store at 2-8°C. Bring reagents to room temperature (15-30°C) before use. Avoid contamination of reagents. Once the foil pouch containing one cassette is opened, it should be used within 12 hours. Assuming good laboratory practices are used, opened reagents remain stable as indicated by the expiration date.

COLLECTION AND HANDLING

ImmunoFLOW Test is performed on serum. Store samples at room temperature for no longer than seventy-two (72) hours. If the assay will not be completed within seventy-two hours, refrigerate the sample at 2-8°C. IgM activity may be affected by freezing the specimen.

PROCEDURE

MATERIALS PROVIDED

Cassette	Color M
Wash 2	Diluent

MATERIALS REQUIRED BUT NOT PROVIDED

Collection apparatus	Timer
Pipette	Test tubes
Control Reagents	Camera (optional)

SET-UP

1. Remove cassette(s) from foil pouch - 1 per test.
2. Remove Diluent, Wash 2 and Color M from kit.
3. Prepare a 1:2 (50:50) dilution of the patient specimen or control using specimen diluent (Diluent). (For example, add 100 µL patient sample into 100 µL Diluent).

ASSAY PROCEDURE

1. Add **one hundred (100) microliters (µL)** of Wash 2 to the cassette. Allow all liquid to flow through device.
2. Add **100 µL** of 1:2 **diluted** patient or control specimen. Allow all liquid to flow through the device. (Note: If sample takes longer than one (1) minute to flow through cassette, do not continue. Attempt run once more. If sample continues to not flow, this sample is not acceptable for procedure. If this commonly occurs, report problem to GenBio. Typically this is caused by highly lipemic sera, but may occur without apparent cause.)
3. Add **100 µL** of Wash 2 to the cassette. Allow all liquid to flow through device.
4. Add **100 µL** of Color M to the cassette. Allow all liquid to flow through device.
5. Add **100 µL** of Wash 2 to the cassette. Allow all liquid to flow through device.
6. Read the results within **two (2) minutes**. (A permanent record may be made using a digital camera.)

INTERPRETATION

Reactive	A red dot is visible
Not Reactive	No dot is visible

It is recommended that users initially test a series of presumptive negative specimens. If needed, GenBio can provide a positive serum control (Part Number 800-3925). Please contact GenBio's Technical Service.

CLINICAL INTERPRETATION

Positive	All dots reactive
Low Positive	Control and dot T reactive
Negative	Only control dot reactive
Do Not Interpret	No dots reactive

For best clinical correlation, if not certain, interpret as negative. In some cases, dot T may report less color intensity than dot 2. The control dot usually exhibits the strongest color intensity.

QUALITY CONTROL

The assay is performed at room temperature (18-27°C).

Testing should be according to guidelines or requirements of local, state, and/or federal regulations or accrediting organizations. Unless otherwise required, it is recommended that control sera be tested upon receipt of a kit.

Each cassette includes a reagent positive control. The background area around the dot is the reagent negative control. The reagent positive control dot must be reactive and the background must be white to pale pink. If the reagent control dot is not reactive or the background is reactive, do not interpret the cassette.

It is recommended an initial assay validation using positive and negative control sera be done before introducing the test into the facility. This is done by testing both negative and positive specimens. It is also recommended that the facility test these negative and positive samples upon receipt of a new shipment.

LIMITATIONS

The values obtained in the assay are intended to be an aid to diagnosis only. Clinical interpretation also requires consideration of the patient's history, physical findings and other diagnostic procedures.

M. pneumoniae complement-fixing antigen may cross-react with other *Mycoplasma* IgM specific antibodies.

Results obtained from immunocompromised individuals should be interpreted with caution.

The performance characteristics have not been established for any matrices other than serum.

There is a possibility of assay cross-reactivity with specimens containing rheumatoid factor.

The prevalence of the analyte will affect the assay's predictive value.

EXPECTED RESULTS

Specific IgM may be present in specimens collected from healthy ("presumptive negative") subjects. Low amounts of specific IgM is present in some healthy subjects. Eighty-three specimens collected in a U.S. asymptomatic adult population were tested using an enzyme-immunoassay (EIA) (ImmunoWELL M. pneumoniae IgM Test) and ImmunoFLOW. One specimen reported EIA positive (1050 units/mL) and two specimens reported EIA low positive (807 and 761 units/mL) and all three reported ImmunoFLOW low positive. The remaining specimens reported EIA negative. One presumptive negative specimen (481 units/mL) was ImmunoFLOW low positive yielding **relative specificity of 99% (80/81)**.

PERFORMANCE

ImmunoWELL Mycoplasma IgM Test uses the same complement-fixing antigen and was validated using case-defined acute and convalescent specimens. Using ImmunoFLOW as reference, specimens were supplied to four sites. Sites A, B and C are located in the European Union. Site D is GenBio. Six replicates of each specimen listed in Table 1 were tested. The same specimens were provided to all sites.

Sensitivity, relative to EIA activity, is 100% (96/96)
(Table 1: Performance Related to IgM Activity).

Reproducibility is 100% (144/144) for specimens reporting values significantly above or below the clinical cutoff region. Two borderline specimens (853 and 644 units/mL) respectively reported 77% and 65% reactivity. Interpretation of borderline reactivity depends on the technician's interpretation. Data presented in Table 1 report results of eight technicians. Interpretation of the borderline specimens ranges from 0-100% reactivity.

Table 1: Performance Related to IgM Activity

Units/mL IgM	Interpretation	Percent Positive or Low Positive
1477	Positive	100% (48/48)
1399	Positive	100% (48/48)
853	Low Positive	77% (37/48)
644	Negative	65% (31/48)
37	Negative	0% (0/48)

BIBLIOGRAPHY

1. **Ali, N H, et al.** The Clinical Spectrum and Diagnosis of Mycoplasma pneumoniae Infections. *Q J Med.* 1986, Vol. 58, p. 227.
2. **Foy, H M.** Mycoplasma Pneumoniae. [book auth.] A S Evans. *Bacterial Infections of Humans: Epidemiology and Control.* New York : Plenum Publishing, 1982, p. 345.
3. **Foy, H M, et al.** Long Term Epidemiology of Infections with Mycoplasma pneumoniae. *J Infect Dis.* 1979, Vol. 139, p. 681.
4. **Cherry, J D, Hurwitz, E S and Welliver, R C.** Mycoplasma pneumoniae Infections and Exanthems. *J Pediatr.* 1975, Vol. 87, p. 369.
5. **Levy, M and Shear, N H.** Mycoplasma pneumoniae Infections and Steven-Johnson Syndrome: Report of Eight Cases and Review of the Literature. *Clin Pediatr.* 1991, Vol. 30, p. 42.
6. **Sillis, M.** The Limitations of IgM Assays in the Serological Diagnosis of Mycoplasma pneumoniae Infections. *J Med Microbiol.* 1990, Vol. 33, p. 253.
7. **US Centers for Disease Control.** *Manual Guide – Safety Management No. CDC-22 Decontamination of Laboratory Sink Drains to Remove Azide Salts.* Atlanta : Centers for Disease Control, 1976.
8. —. *HHS Publication No. (CDC) 93-8395, 3rd ed: Biosafety in Microbiological and Biomedical Laboratories.* Washington DC : US Government Printing Office, 1993.
9. **US Department of Labor, Occupational Safety and Health Administration.** *29 CFR Part 1910.1030, Occupational safety and health standards, bloodborne pathogens.*
10. **US Department of Health and Human Services.** *HHS Publication No. (CDC) 21-11: Biosafety in Microbiological and Biomedical Laboratories. 5th ed.* Washington DC : US Government Printing Office, 2009.
11. **World Health Organization.** *Laboratory Biosafety Manual 3rd ed.* Geneva : World Health Organization, 1991.

To place an order for ImmunoFLOW products, contact your local distributor or call GenBio directly for the distributor nearest you and for additional product information.

For assistance, please call toll-free 800-288-4368.



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